

Gallios Training

Beckman Coulter Company Guide

https://www.beckmancoulter.com/ucm/idc/groups/public/documents/webasset/glb_bci_150264.pdf

Calendar for time sign up:

<http://bidflow.calendarhost.com/>

Sheath & Waste Tanks

- Please check before starting
- External Tanks
 - Will beep/ notify if empty or full
- Reservoirs
 - Blue reservoir is cleansing solution

Power Supply

- Circuit breaker → on and off switch (hard re-set)

Configuration box

- Can change filters/ configurations
- Can take out filters/ swap around
- If you have to change check with someone in lab

Drip Chamber

- If running for 10+ minutes / sample then drip chamber must empty before next sample

Carousel

- Can set up tubes & leave
- Samples 400-500 ml volume per sample recommended for running (minimum 200 mL)

Kaluza - Acquisition (software)

- Turning machine on/ off done via software
- Blue Orb- Acquisition (use to turn on)
- Red Orb- Analysis portion
 - Can analyze while samples are running

Fluorescence Channels

<https://www.bsf.star.edu.sg/Ads%20Pictures/Gallios%20Filters%20Config.pdf>

Main Menu

1. FL1- FITC
 2. FL2 – PE
 3. FL3 -ECD/ PE Texas red/ PI
 4. FL4 -PE CY5
 5. FL5- PE CY7
 6. FL6 –APC
 7. FL7 -A700
 8. FL8 – APC CY7
 9. FL9 – Pacific Blue
 10. FL10 – Pacific Orange
- Instrument Control → Initialize → Cycles up instrument
 - Clean → cleans lines (blue solution)
 - Open
 - New protocol
 - Import old protocol
 - Can choose to enable all detectors OR pick corresponding
 - EDIT work list → edits carousel tubes
 - Duplicate (links to protocol)
 - Name tubes/ link to specific protocol → Done editing
 - Clear work list → resets/ deletes
 - Choose output location → Users file → Make folder

Instrument Controls

- Acquire single (acquires one)
- Acquire → runs through entire sample
- Prime 3x → clears lines/ clogs
- Set up mode (blue) → acquires *without saving*
 - Use for setting up voltages, compensation
 - Dictate # events

Acquisition Controls

- Choose number of events to accumulate OR time
- Compensation – sliders
- Voltage – sliders
- Speeds → low, med high

Plots & Gates

- Histograms, Plots, Gates
- Add all plots (option)
 - Can set up to preference
 - Additional plots

Running samples

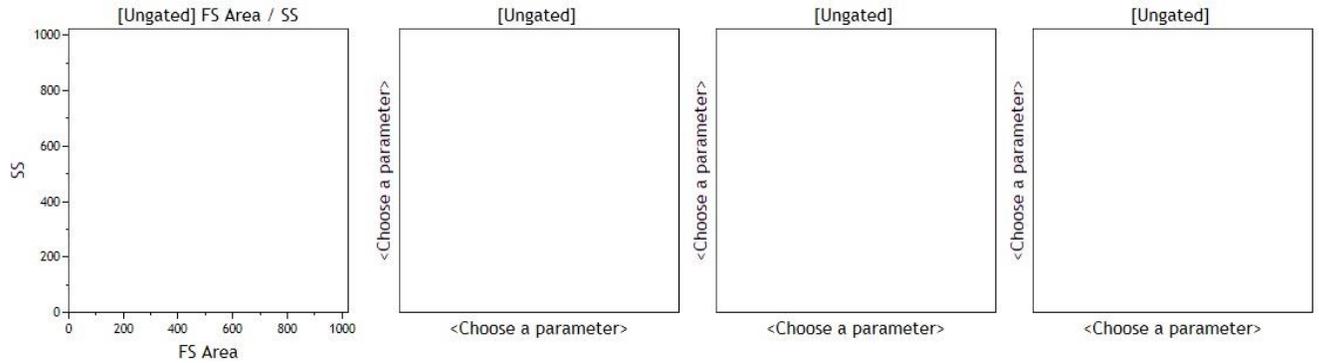
- Right click → Radial Menu → brings up options
- Features → Undo/ Redo

Hardware

- Discriminator → “Threshold”
 - Set to 100 by default→ can adjust based on sample
- Particle Size
 - Submicron: microparticles
 - Small: Basic whole blood components (3micron-20micron)
 - Large: < 20 micron
- Voltage → fine adjustment
- Gain → course adjustment

Starting Experiment

- Access Kaluza (blue orb) and load tubes accordingly
- Open
 - New protocol
 - Import old protocol
- Choose Proper Band Passes:
 1. FL1- FITC
 2. FL2 – PE
 3. FL3 -ECD/ PE Texas red/ PI
 4. FL4 -PE CY5
 5. FL5- PE CY7
 6. FL6 –APC
 7. FL7 -A700
 8. FL8 – APC CY7
 9. FL9 – Pacific Blue
 10. FL10 – Pacific Orange
- Instrument Control →Initialize → Cycles up instrument
- Prime 3x
- Enter Set up mode
 - # of events, amount of time, and speed of acquiring
 - Determine Threshold and particle size wanted
 - Create graphs and plots wanted for recording data
 - Basic Example:



- Acquire Single on unstained sample: KEEP SET UP MODE ON
 - Adjust Gain → then adjust voltages for FSC & SSC
 - Adjust Fluorescence channels
 - Rename graphs
 - Turn off set up to record data and run remaining sample
- Acquire Single on compensation controls: SET UP MODE
 - Adjust Gain → then adjust voltages for FSC & SSC
 - Adjust Fluorescence channels
 - Rename graphs
 - Can also type in via compensation matrix
 - Turn off set up to record data and run remaining sample
- Repeat above step for each control
- Acquire for rest of mixed samples
- Use set up modes to run through compensation controls
- Can save worklist & save compensation
- Hit Red Orb to go to Kaluza Analyzation software

Kaluza Analyzis (Red Orb)

- New Composite: can select all files → new composite → merges data sets
- Plots & Gates: new options from acquisition
 - Overlay plots
 - Comparisons